

Taq Polymerase Enzyme (5U/μl)

Catalog	Description
HL - Taq - 50	250Units
HL - Taq - 100	500Units
HL - Taq - 200	1000Units

Store at - 20°C
 PI/HL - Taq - 01

Product Description

Taq Polymerase Enzyme is a temperature-resistant DNA-dependent polymerase used for the amplification of DNA using Polymerase chain reaction (PCR). It is a thermostable DNA polymerase that has 5'→3' polymerase and exonuclease activity but lacks 3'→5' exonuclease (proofreading) activity. The enzyme is ideal for gene amplifications up to 3kb.

Source

Modified synthetic *Taq* DNA Polymerase gene from *Thermus aquaticus* is expressed and purified in *E. coli* strain.

Reagents Supplied

- *Taq* Polymerase Enzyme (5U/μl),
- 10X *Taq* buffer with MgCl₂ (1ml)
- 25mM Magnesium Chloride (500μl)

Unit definition (5U/μl)

One unit is the amount of enzyme required to catalyze the incorporation of 10 nmol of dNTP into acid-insoluble material in 30 minutes at 65°C

Storage conditions

The recommended storage condition is -20°C.

PCR Protocol

Component	Volume (μL)
10X <i>Taq</i> Buffer with MgCl ₂ (10X)	2.5
10 mM dNTPs	0.5
Forward primer (10 pmol/μL)	0.4-0.7
Reverse primer (10 pmol/μL)	0.4-0.7
Template DNA	Variable (user defined)
<i>Taq</i> Polymerase Enzyme (5U/μL)	0.5
Molecular Grade Water	Make up to 25

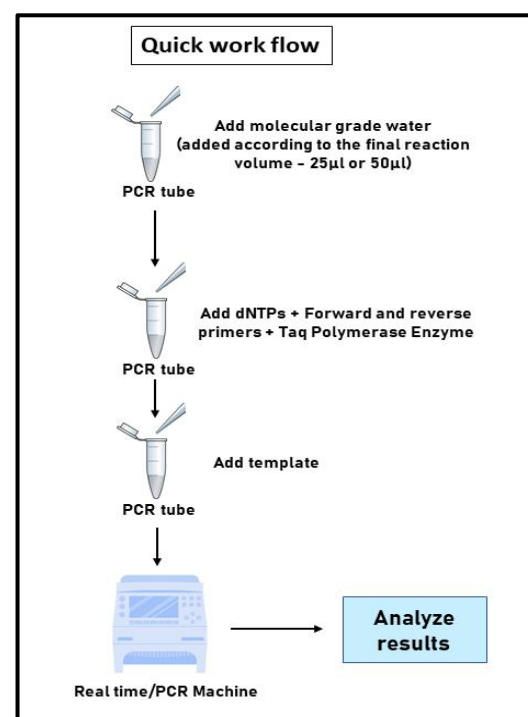
Note: In the above reaction, the concentration of MgCl₂ will be 2.5mM and this can be adjusted according to the experiment with the additional MgCl₂ provided if required

Recommended PCR Program

Operation	Temp	Time	Cycles
Initial denaturation	95°C	1-5 min	1
Denaturation	95°C	30 sec	30-35 cycles
Annealing	45-68°C (User defined)	30-60 sec	
Extension	72°C	1 min/kb	
Final Extension	72°C	5-10 min	1

Notes:

Mg⁺² ions: *Taq* DNA polymerase effectively amplifies the target with 1.5 mM MgCl₂. However, the reactions can be optimized for DNA samples which may require higher concentrations of MgCl₂. Very high levels of Mg⁺² promote non-specificity while low concentration of Mg⁺² may reduce the yield.



Reference Guide

Additive: The presence of the inhibitors in the samples can affect PCR. Using reagents like DMSO, betaine, etc. can improve the PCR outcome. However, this should be done with careful standardization

Template: Optimal DNA template concentration usually used in PCR is up to 1 ng for both plasmid and phage DNA while 10-15ng for genomic DNA. The higher concentrations of template DNA generate non-specific PCR products whereas lower concentrations affect the PCR amplification

dNTPs: Generally, 200 μ M of each dNTP is recommended for the PCR. However, on a few occasions, higher concentrations of dNTP may be required whereas the concentration of Mg⁺² has to be adjusted accordingly as Mg⁺² binds to dNTPs.

Primers: The suggested concentration of primers for PCR ranges between 0.1 and 1 μ M.

Applications

- Primer extension
- Routine PCR amplification for TA cloning
- DNA sequencing
- RFLP

***We recommend referring to our quick enzyme guide to choose the right enzyme for your needs**

Taq Polymerase Enzyme	QuickStart Taq Polymerase Enzyme	HotStart Taq Polymerase Enzyme	Klen BST Polymerase Enzyme
Used for endpoint PCR	Real-time PCR and Endpoint PCR	Real-time PCR	Real-time LAMP and LAMP
Works with temperature shifts	Works with temperature shifts	Works with temperature shifts	Ideal for isothermal PCRs 60°C – 65°C
No activation	30 seconds activation	15 minutes activation	No activation

Other PCR products from Huwel LifeSciences that you may be interested

S.No.	Product description	Catalogue No.
1.	Universal PCR Mix (2X)	HL – UPM – 100 – 1.25ml
2.	UltraFast Real Time PCR Mix 5X	HL – UFPCM – 100 – 500 μ l HL – UFPCM – 200 – 1ml
3	UltraFast Real Time PCR Mix 5X with UDG	HL – UUFPCM – 100 – 500 μ l HL – UUFPCM – 200 – 1ml
4	HotStart Taq Polymerase Enzyme (5U/ μ l)	HL – HSTaq – 50 – 250Units HL – HSTaq – 100 – 500Units HL – HSTaq – 200 – 1000Units
5	QuickStart Taq Polymerase Enzyme (5U/ μ l)	HL – QSTaq – 50 – 250Units HL – QSTaq – 100 – 500Units HL – QSTaq – 1000Units

References

- Chien, A., Edgar, D.B. and Trela, J.M. (1976) *J. Bact.*, 127, 1550-1557
- Kaledin, A.S., Sliusarenko, A.G. and Gorodetskii, S.I. (1980) *Biokhimiya*, 45, 644-651
- Lawyer, F.C. et al. (1993) *PCR Methods and Appl.*, 2, 275-287
- Longley, M.J., Bennett, S.E. and Mosbaugh D.W. (1990) *Nucleic Acids Res.*, 18, 7317-7322

For further information on protocols and details, please contact our technical support:

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