

Quantiplus® Nipah Virus Detection Kit V2 (Real-Time Qualitative PCR Kit)



QL-NIV2-25 : 25 rxns
 QL-NIV2-50 : 50 rxns
 QL-NIV2-100 : 100 rxns

RUO

PI/QLNIV2-00

Intended Use:

Quantiplus® Nipah Virus Detection Kit is a Real-Time PCR based in vitro diagnostic assay for detection of Nipah virus in throat swabs in VTM (Viral transport medium), Plasma, CSF and Urine. The detection is based on the amplification of a specific region within the nucleocapsid protein gene of the Nipah virus. The kit consists of amplification mix, Primers and Probe mix for Nipah virus and Internal Control and also Nipah Positive control. The kit simultaneously detects the nucleocapsid gene of the Nipah virus and the human endogenous gene RNase P. The RNase P serves as an endogenous internal control and helps to monitor the nucleic acid integrity and amplification. However, RNase P amplification may or may not be detected for CSF and urine sample types.

Background Information:

Nipah virus belongs to the family *Paramyxoviridae*, genus *Henipavirus*. It is a zoonotic virus, spreading between animals and humans. Fruit bats (genus *Pteropus*) is the animal host reservoir for the virus. Infected fruit bats can spread the disease to humans and pigs. People get infected if they have close contact with an infected animal/human or its body fluids. Infections can range from asymptomatic infection to acute respiratory infection (mild, severe), and fatal encephalitis. The case fatality rate is estimated at 40% to 75% based on the outbreaks between 1998-2018. Nipah Virus infection can be very difficult to diagnose owing to the non-specific early symptoms of the illness. However, early detection and diagnosis helps to increase chances of survival among infected individuals, to prevent transmission to other people, and to manage outbreaks.

Kit Components

Color Coding (Caps)	Contents	Description	25 rxns (QL-NIV2-25)	50 rxns (QL-NIV2-50)	100 rxns (QL-NIV2-100)
Green	R Fast Core qPCR Mix (2X)	Amplification Reagent	1 x 325 µL	1 x 650 µL	2 x 650 µL
Amber	Nipah Primer Probe Mix	Nipah, Internal Control Primers and Probes Mix	1 x 50 µL	1 x 100 µL	2 x 100 µL
Red	Nipah Positive Control	Nipah Virus Positive control	1 x 100 µL	1 x 100 µL	2 x 100 µL
White	Huwel PW	Purified water	1 x 500 µL	1 x 500 µL	2 x 500 µL

Note: Please pay attention to the cap color coding and the tube contents.

Storage and Transportation Conditions:

The kits should be transported at temperature below $-20\text{ }^{\circ}\text{C}$. The kit is stable until the expiry date mentioned on the package, if the storage temperature is within $-20 \pm 5\text{ }^{\circ}\text{C}$. More than 4X freezing and thawing cycles reduces the assay sensitivity. For intermittent usage the reagents should be frozen in aliquots.

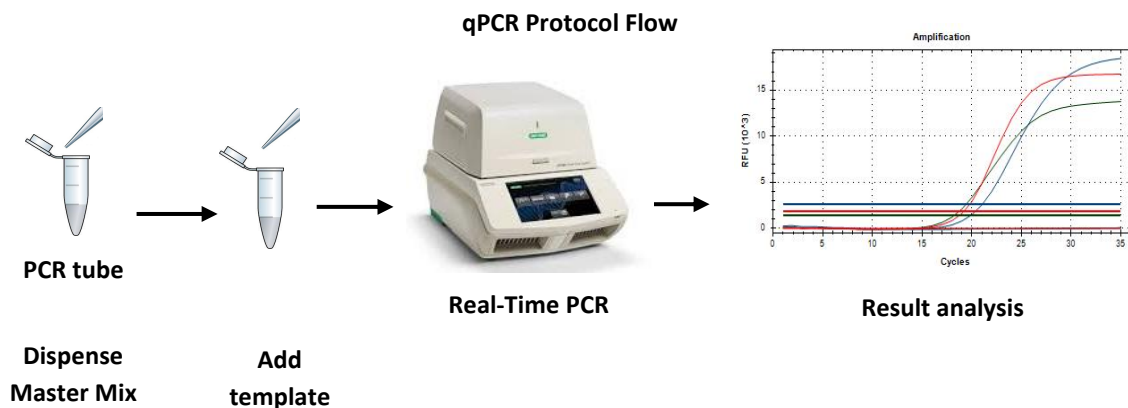
Assay Procedure:

RNA Extraction

Quantiplus® Nipah Virus Detection Kit (Real-Time Qualitative PCR Kit) has been validated using the following Viral RNA extraction kits are as follows:

S. No.	Name of the Extraction Kit	Recommended Sample volume for Extraction	Recommended Final Elution volume
1.	Huwel Nucleic Acid Extraction Kit - Version 2.0 (Cat. No. HL-NAX-100)	200 μL	100 μL
2.	QIAamp® Viral RNA Mini Kit (Cat. No. 52904)	140 μL	50 μL

Note: Customer can also validate their own extraction process using other viral DNA extraction Kits.



Preparation of Reaction Master mix

Components	Volume per reaction (for 25 μL)
R Fast Core qPCR Mix (2X)	13.0
Nipah Primer-Probe Mix	2.0
Extracted RNA/ Nipah Positive Control /Huwel PW	10.0

It is necessary to keep all components at $+2\text{ }^{\circ}\text{C}$ to $+8\text{ }^{\circ}\text{C}$ during the PCR preparation. Close the tubes and centrifuge briefly before proceeding to the thermal cycler.

Cycling Conditions:

Steps	No. of cycles	Temperature (°C)	Time
1. (Reverse Transcription)	1	53	5 min
2. (Initial denaturation)	1	95	3 min.
3. (PCR cycling)	45	95	10 sec.
		60*	30 sec.
*Plate read/Data acquisition in FAM and TEXAS RED channels.			

Sample analysis and Interpretation

Interpret the values for unknown samples would appear in the result column in FAM Channel. Samples showing no amplification in FAM channel should show amplification in Texas Red channel, and then only results should be considered. The negative control should not show any value in the FAM and Texas Red channel. It is important to analyze the target channels (FAM and Texas Red) individually in separate windows for better clarity.

S.No	FAM (Nipah)	TEXAS RED(IC)	Fluorophore		Conclusion
			Interpretation		
1	√	√	Nipah RNA detected	←	Proceed for further Analysis
2	√	-			
3	-	√			
4	-	-	Possible inhibition of PCR		Dilute the RNA sample (1:10) and repeat the Assay

Note: All the Target channels (FAM and Texas Red) to be analyzed individually.

Validated Instruments

- Thermo QS5 Real-Time PCR System
- Bio-Rad™ CFX 96



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